

Figure 3:
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f)
composition of *Paramecium*, *Gracilaria*, *Trichocapsa*, and *Pennakia* from
each sampling site and for all sites combined (g,h). The different sampling
methods are distinguished (shallow, shallow stream, deep stream).
The size distribution of all species caught (i) is presented by sampling
date.
No shallow or deep stream samples were collected during May. No pennakia
samples were collected at the F&B site during September. Only the OUTLET
site was sampled during November.

Size distribution

May
June
July
September
November

Figure 7:

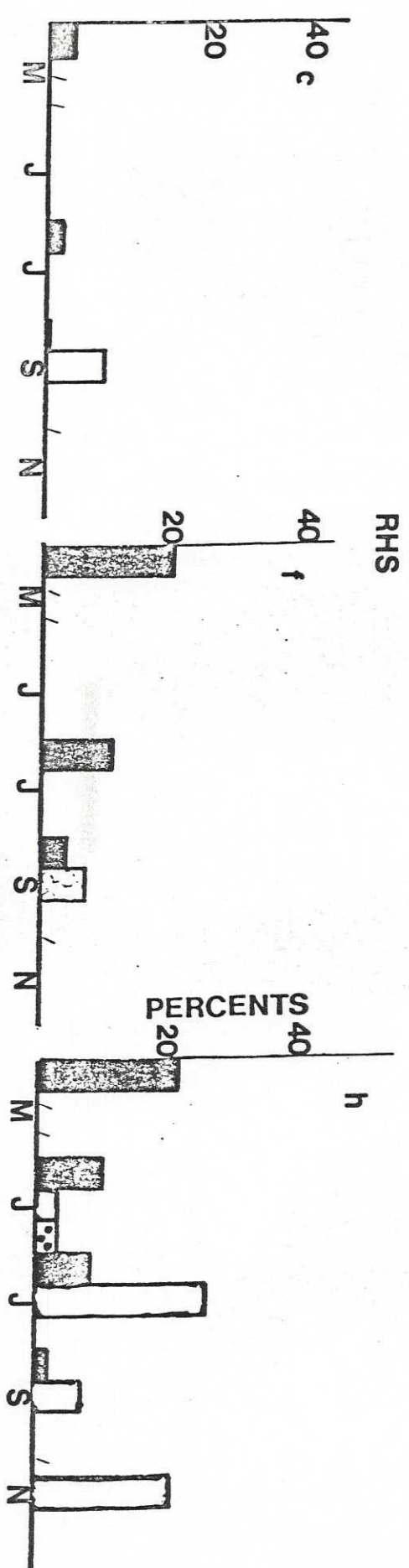
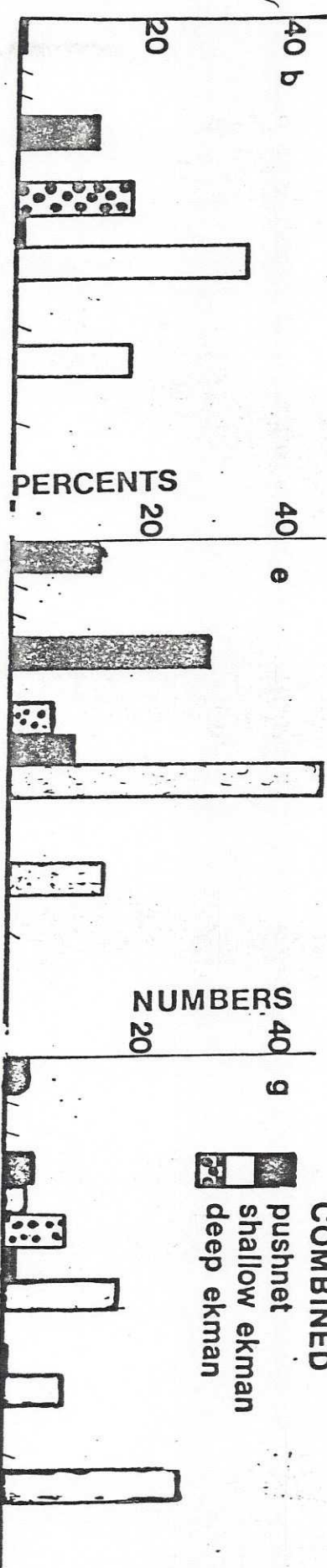
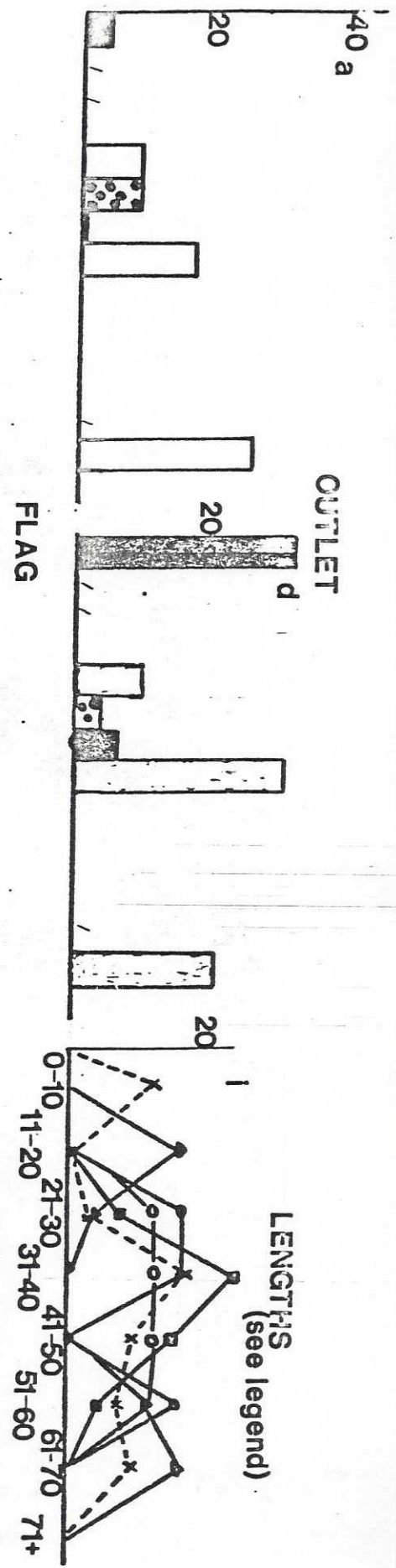
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of Decapods, Orconectes rusticus, to benthic samples from each sampling site and for all sites combined (g,h). The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman). The size distribution of all crayfish caught (i) is presented by sampling date.

No shallow or deep ekman samples were collected during May. No pushnet samples were collected at the FLAG site during September. Only the OUTLET site was sampled during November.

Size distribution



NUMBERS PER METER SQUARED



DECAPODA

DECAPODA



NUMBERS PER METER SQUARED

deep shrimp
shallow shrimp
combined

LENGTHS
(see legend)

The Decapoda include the largest individuals caught as benthos and were a monospecific assemblage of Orconectes rusticus, ranging in size from 10-75 mm (see Figure 7). Crayfish tend to be scavengers (Crocker and Barr, 1971). A qualitative sample of 5 crayfish stomachs from the July collections confirmed that the crayfish were consuming chironomid larvae and detritus.

Figure 7 shows the numbers of crayfish caught in benthic samples and the percent volume they contributed to respective samples. In July, the numbers at the FLAG ekman site reached 34 individuals per meter squared contributing 45 percent of the sample volume. The large size range of crayfish indicates that the crayfish may have a three year life cycle in the lake (Crocker and Barr 1971).

Order Odonata

Anisoptera and Zygoptera nymphs were distinguished because of their different body morphologies.

Suborder Anisoptera:

The dragonfly nymphs in Sunfish Lake proper are all of the family Lebullidae although in the OUTLET stream itself rare catches of other families were made. Figure

The *Desmophis* include the largest individuals caught on beaches and were a monospecific assemblage of *Desmophis* insects, ranging in size from 18-25 mm (see Figure 1). Crayfish tend to be scavengers (Crocker and Burt, 1971). A qualitative sample of 5 crayfish from the July collection indicated that the crayfish were consuming filamentous larvae and detritus.

Figure 1 shows the numbers of crayfish caught in beach samples and the percent volume they contributed to respective samples. In July, the numbers at the RAB (mean size reached 35 individuals per meter) were not contributing 45 percent of the sample volume. The large size range of crayfish indicates that the crayfish may have a three year life cycle in the lake (Crocker and Burt 1971).

Order Odonata

Anisoptera and Zygoptera nymphs were distinguished because of their different body morphologies.

Suborder Anisoptera

The dragonfly nymphs in South Lake proper are all of the family Libellulidae although in the outlet stream Libellulidae and other families were noted. Figure

8 diagrams the numerical abundances and percent volumes of the *Anisoptera* nymphs on a seasonal basis. Also shown is a breakdown of the counts into two size categories, those less than 15mm in length and those greater than 15mm in length. 15mm is a natural break in the size distribution that probably corresponds to one of the instar moults. Wing pads were only visible on nymphs greater than 15 mm in length. The largest Lebulidae caught were 23mm in length and the smallest were 6mm. It can be seen from the single size division that, although a few, large *Anisoptera* nymphs are found in May, the preponderance of the June collections are of small individuals. In July the emphasis shifts to larger individuals and this trend persists into September when there is a reappearance of large numbers of smaller individuals contributing to the high, 260 individuals per meter squared, numerical abundance composing 22 percent of the volume in surface ekman samples. This suggests a seasonal cycle of emergence in May, June and July and a recruitment of early instar nymphs in September. November localization is in the shallow water. The *Anisoptera* nymphs were found in association with the *Chara*. This probably relates to the concentration of food in that area and to the protection offered by the cover as the nymphs are generally slow moving (Walker, 1953). Lebullidae are predatorial (Merritt and Cummins 1978).

8 diagrams the numerical abundances and percent volumes of the antipodal nymphs on a seasonal basis. Also shown is a breakdown of the counts into two size categories, those less than 15mm in length and those greater than 15mm in length. 15mm is a natural break in the size distribution that probably corresponds to one of the instar periods as wing pads were only visible on nymphs greater than 15 mm in length. The largest leafhoppers caught were 23mm in length and the smallest were 6mm. It can be seen from the single size division that, although a few, large antipodal nymphs are found in May, the preponderance of the June collections are of small individuals. In July the emphasis shifts to larger individuals and this trend persists into September when there is a reappearance of large numbers of smaller individuals contributing to the high 38% individuals per meter squared, numerical abundance composing 12 percent of the volume in surface areas sampled. This suggests a seasonal cycle of emergence in May, June and July and a recruitment of early instar nymphs in September. November localization is in the shallow water. The antipodal nymphs were found in association with the Chara. This probably relates to the concentration of food in that area and to the protection offered by the cover as the nymphs are generally slow moving (Walker, 1951). Leafhoppers are predators (Merritt and Cummins 1978).

Figure 8:
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f)
contributions of *Antipathera myriophylla* (Labridae) to benthic samples from
each sampling site and for all sites combined (g,h). The different sampling
methods are distinguished (bottom, shallow water, deep water).
Seasonal changes in the numbers of individual species < 15 cm in size (i,j)
are also shown.
In shallow or deep water samples were collected during May. In bottom
samples were collected at the TMS site during September. Only the GERT
site was sampled in November.

Figure 8:

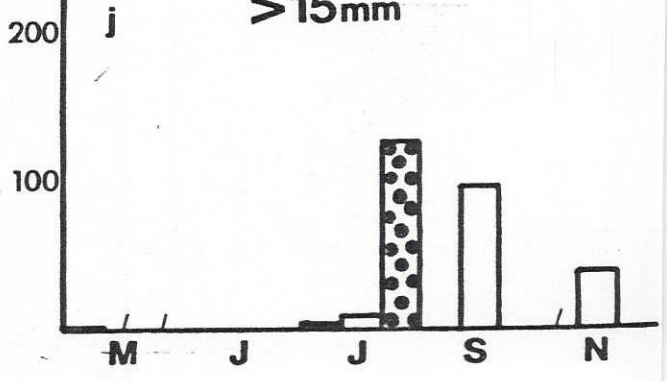
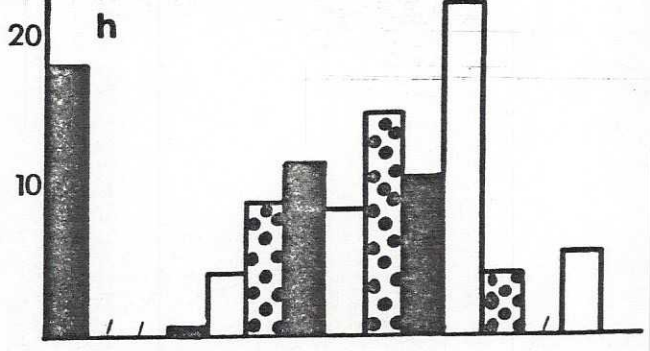
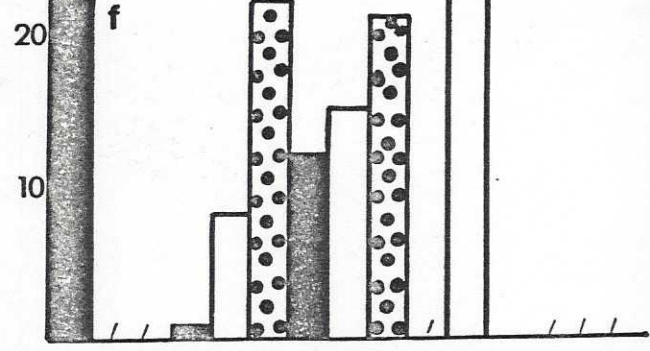
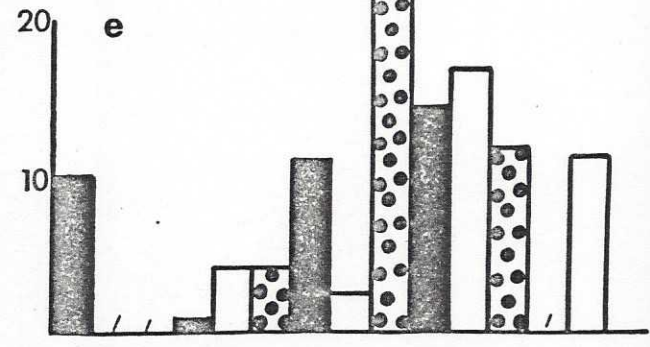
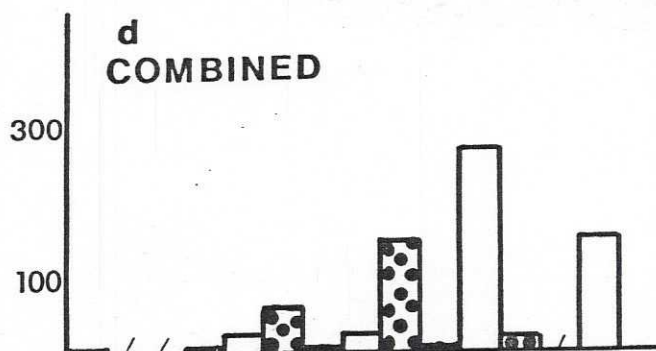
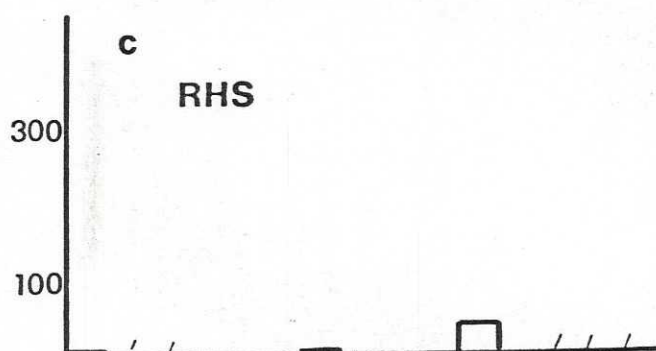
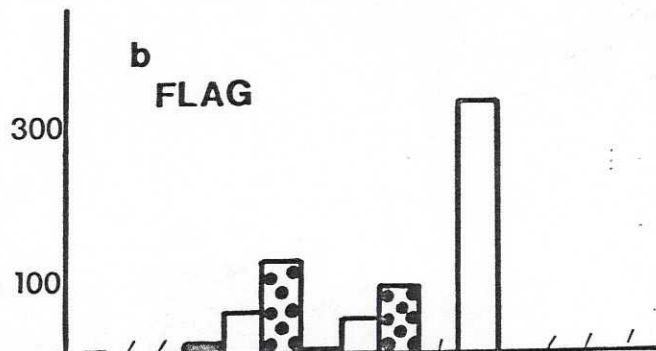
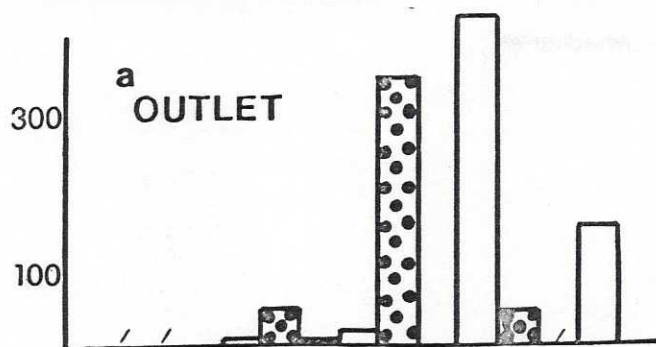
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of Anisoptera nymphs (Lebullidae) to benthic samples from each sampling site and for all sites combined (g,h). The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman). Seasonal changes in the numbers of individual nymphs <15 mm or >15 mm (i,j) are also shown.

No shallow or deep ekman samples were collected during May. No pushnet samples were collected at the FLAG site during September. Only the OUTLET site was sampled in November.

ANISOPTERA

pushnet
shallow ekman
deep ekman

NUMBERS PER METER SQUARED



Suborder Zygoptera:

The seasonal changes in abundance and percent volume of the damselfly nymphs are shown in figure 9. A single genus, Protallagma, was represented (Family Coenagrionidae). Low numbers of nymphs present during May sampling account for almost 5 percent of the sampled biomass. This declines to less than 2.5% of the volume in June and July with damselfly nymphs absent at the OUTLET site. This indicates an emergence which is followed in September by a large recruitment raising the abundance to over 120 individuals per meter squared. This 10 fold numerical increase over the July sample reflects a 5 fold increase in the percent volume contribution of the nymphs. Recorded lengths ranged from 6 to 12 mm exclusive of caudal gill filaments with larger individuals recorded in May and in November and a preponderance of smaller individuals in September. The distribution of the Zygoptera nymphs was similar to that of the Anisoptera nymphs although, wherever found, the numbers of Anisoptera and their percent volume contribution exceeds that of the Zygoptera.

Order Ephemeroptera:

Ephemeroptera, or mayfly nymphs are collector-

The seasonal changes in abundance and percent volume of the damselfly nymphs are shown in Figure 2. A single genus, *Pseudagrion*, was represented (family Coenagrionidae). Low numbers of nymphs present during dry sampling account for almost 5 percent of the sampled biomass. This declines to less than 1.5% of the volume in June and July with damselfly nymphs absent at the outlet site. This indicates emergence which is followed in September by a large resurgence relating the abundance to over 150 individuals per meter squared. This is followed (unpublished) increase over the July sample reflects a 5 fold increase in the percent volume contribution of the nymphs. Recorded lengths ranged from 12 to 15 mm exclusive of caudal gill filaments with larger individuals recorded in May and in November and a predominance of smaller individuals in September. The contribution of the hypogastric nymphs was similar to that of the damselfly nymphs although, however found, the number of Antopoda and their percent volume contribution exceeds that of the Hypogastric.

Order: Hymenoptera

Hymenoptera, or mainly nymphs are collected

Figure 5:

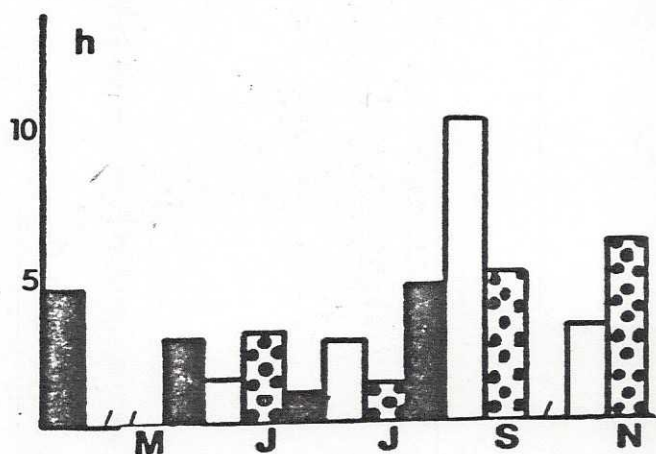
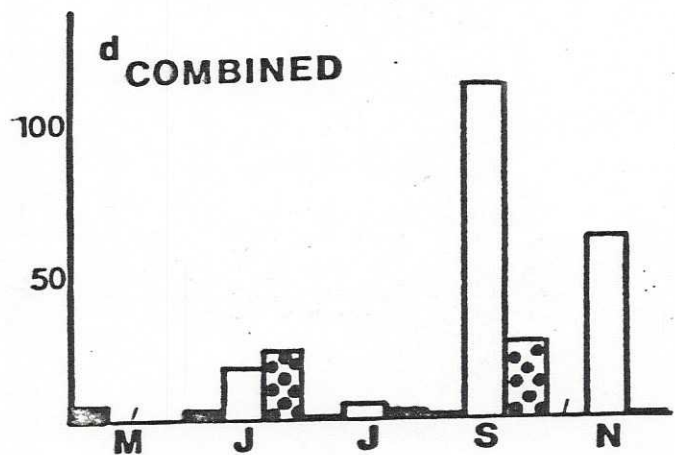
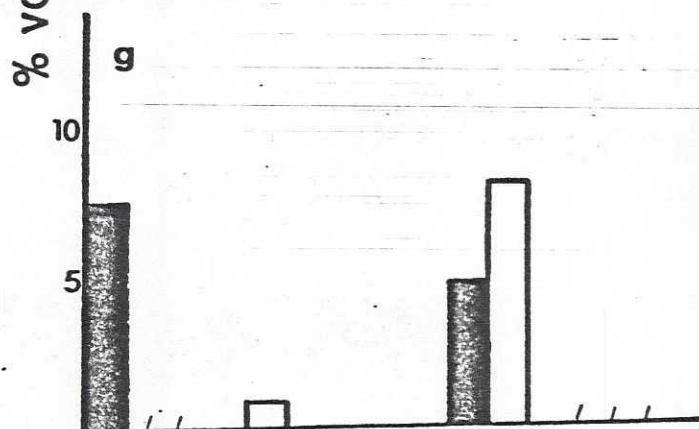
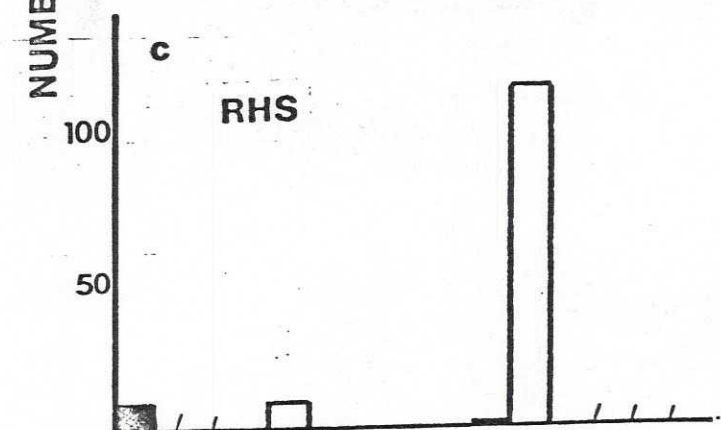
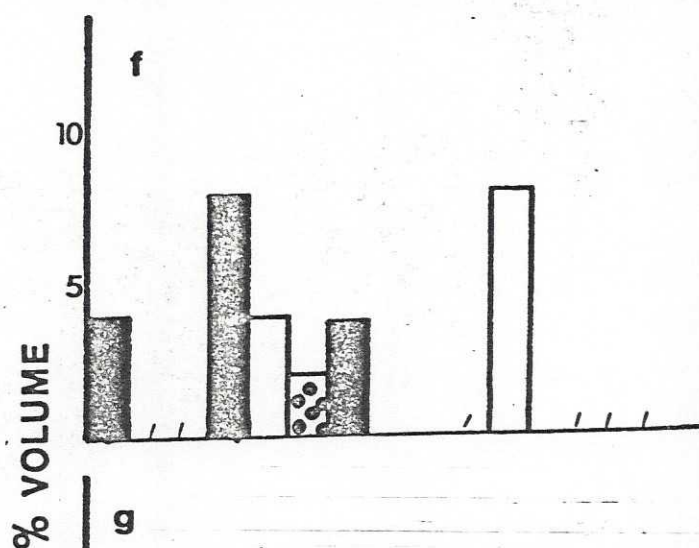
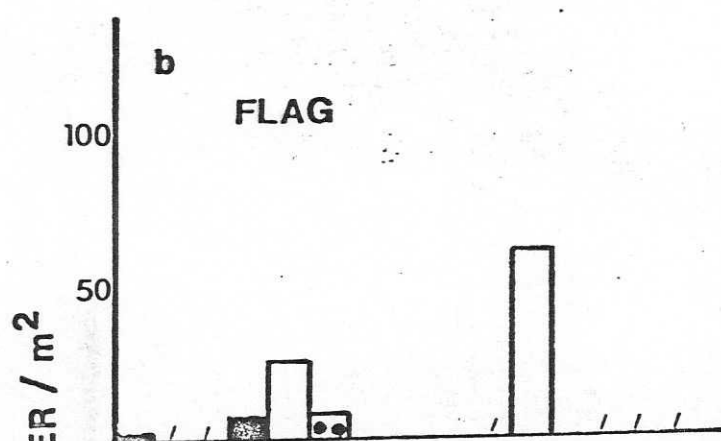
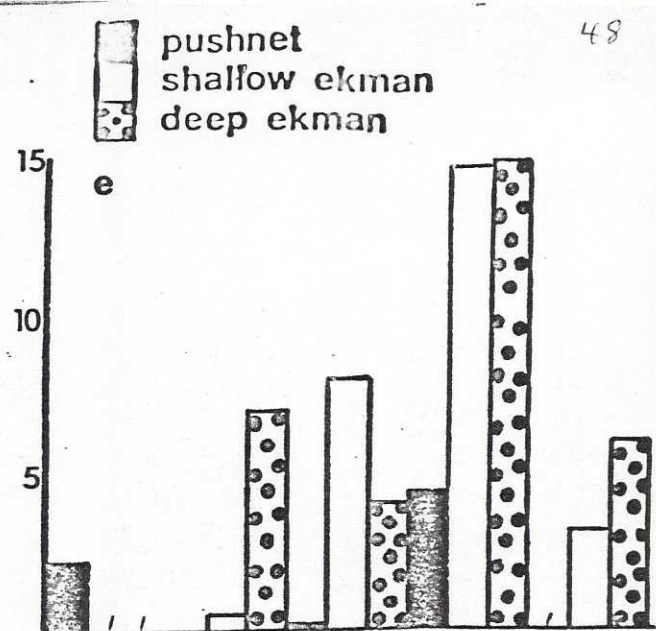
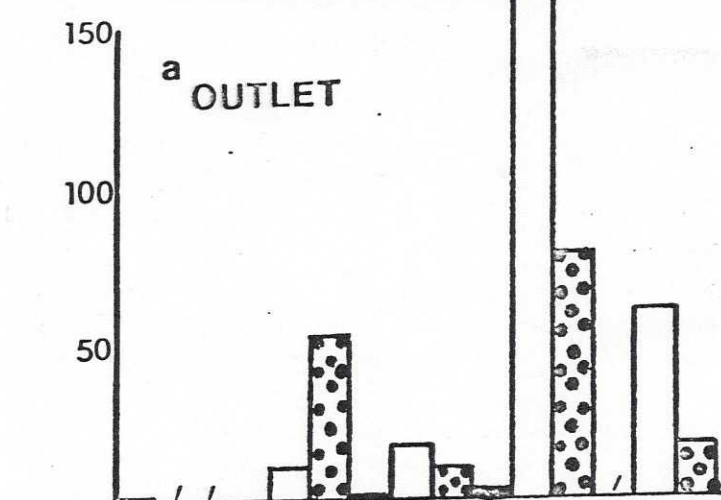
Seasonal changes in the numbers (a, b, c) and the percent volume (d, e, f) contribution of *Epiphyas* nymphs, *Protoparce*, to percent samples from each sampling site and for all sites combined (g, h). The different sampling methods are distinguished (posterior, shallow stream, deep stream). No samples or data shown sampled were collected during May. No samples were collected at the PAC site during September. Only the GUTS site was sampled during November.

Figure 9:

Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of Zygoptera nymphs, Protallagma, to benthic samples from each sampling site and for all sites combined (g,h). The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman).

No shallow or deep ekman samples were collected during May. No pushnet samples were collected at the FLAG site during September. Only the OUTLET site was sampled during November.

ZYGOPTERA



ZYGOTERA

OUTLET



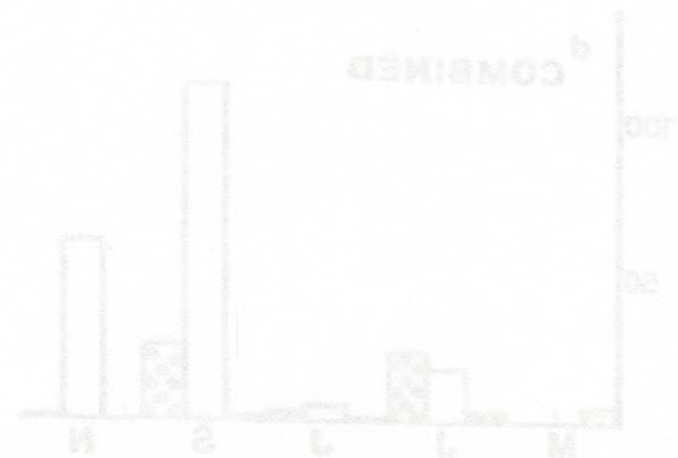
FLAG



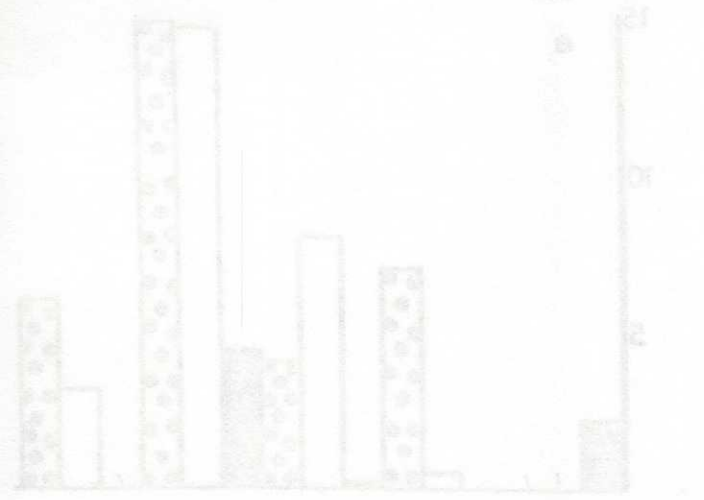
RHS



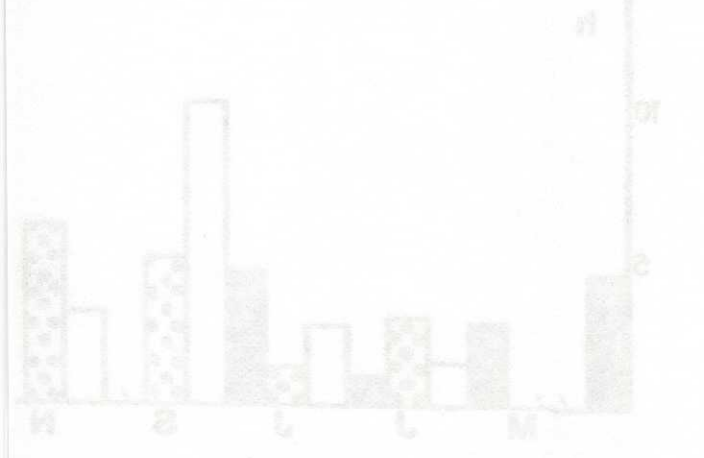
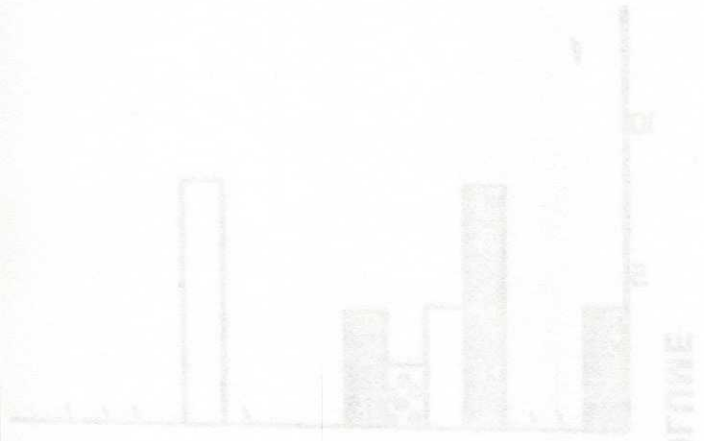
COMBINED



shallow stream
deep stream



COUNT



gatherers rather than predators (Merritt and Cummins, 1978) and are seasonal residents of the benthos. The data for the Order ~~are~~ presented in two distinct groups.

Family Ephemeridae:

The first group is Family Ephemeridae, a monospecific assemblage of Hexagenia bilineata (see Figure 10). The nymphs are large, 16 - 24 mm exclusive of caudal filaments. Although numerically rather sparse they account for a considerable proportion of the sample biomass volume in May and June samples, making the most significant contribution at the relatively impoverished RHS site. Similar to the Odonata nymphs, the OUTLET site is characterized by high numerical densities in July when the nymphs are absent for the other sites. Emergence is complete by the September sampling and there is no evidence for an autumn recruitment of nymphs. This is in accord with the diapause postulated for this species in temperate climates (Edmunds et al, 1971). Barber and Kevern (1973), describe a positive correlation between this species distribution and aquatic macrophytes. The distribution observed in Sunfish Lake was not completely Chara dependent, particularly at the RHS site. This is possibly the result of the predator protection afforded by the semi-

gathered rather than predators (Muller and Gumbel, 1976) and are seasonal residents of the beach. The data for the G-Set are presented in two distinct groups.

Family Ephemeroidea

The first group is Family Ephemeroidea, a monospecific assemblage of *Hexagenia bilineata* (see Figure 10). The nymphs are large, 15 - 25 mm exclusive of caudal filaments. Although numerically rather sparse they account for a considerable proportion of the sample biomass volume in May and June samples, making the most significant contribution at the relatively impoverished K&S site. Similar to the G-Set nymphs, the OUTLET site is characterized by high numerical densities in July when the nymphs are absent for the other sites. Emergence is complete by the September sampling and there is no evidence for an autumn recolonization of nymphs. This is in accord with the diapause period for this species in temperate climates (Edmondson et al., 1971). Rather and Kevan (1973) describe a positive correlation between this species distribution and aquatic macrophytes. The distribution observed in English Lake was not completely *Carex* dependent, particularly at the K&S site. This is possibly the result of the predator protection afforded by the semi-

Figure 10:
Seasonal changes in the numbers (a, b, c) and the percent volume (d, e, f)
contribution of *Isopoda* (Isopoda) groups to benthic
samples from each of the sites and for all the sites combined (g, h). The
different sampling methods are distinguished (pushnet, shallow drag, deep
drag).
During May, shallow and deep drag samples were not collected. No pushnet
samples were collected at the P16C site in September. Only the GUTIER site
was sampled during November.

Figure 10:

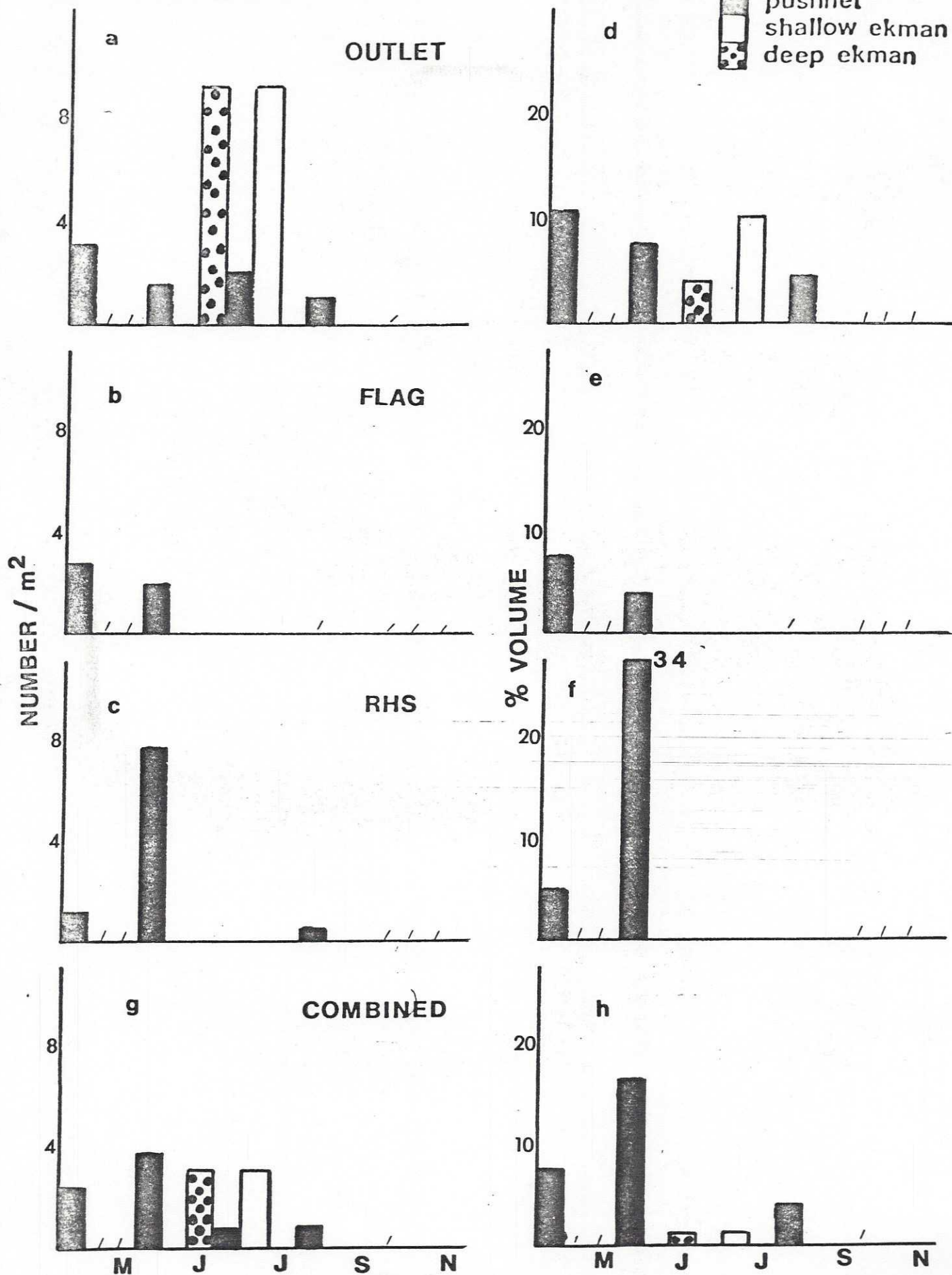
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of Hexagenia bilineata (Ephemeroptera) nymphs to benthic samples from each of the sites and for all the sites combined (g,h). The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman).

During May, shallow and deep ekman samples were not collected. No pushnet sample was collected at the FLAG site in September. Only the OUTLET site was sampled during November.

HEXAGENIA

50

pushnet
shallow ekman
deep ekman



permanent burrows individuals occupy in the substrate (Edmunds et al 1971). Larger bodied individuals tend to be associated with larger grain size sediments (Barber and Kevern, 1973), such as the marl of the RHS site.

Family Ephemerellidae:

The second group of Ephemeropterans are much smaller than the Hexagenia and belong to the genus Ephemerella. They are small, 2 - 6mm, dorsoventrally flattened and tended to be found in association with patchy Chara. Their small size means that the early instars are likely to be underrepresented in the samples. Numerical abundances reach maxima of 400 individuals per meter squared at RHS in June and 450 individuals per meter squared at OUTLET in July accounting for 19 and 26% of the sample volumes respectively. Overall values are considerably lower, approximately 6 percent of the sample volume, but indicate that the nymphs are present throughout the summer and are probably a di- or tri-voltine population of overlapping generations. (Edmunds et al 1971). The Ephemerella, too, are nonpredacious collector-gatherers (Merritt and Cummins 1978).

Order Trichoptera:

The Trichoptera, or caddisflies, are a

potent burrow individuals occupy in the substrate
thousands at a time. Larger bodied individuals tend to
be associated with larger grain size sediments (Barber
and Raven, 1973), such as the wall of the RMS site.

Family Ephemeroidea:

The second group of Ephemeroidea are much smaller than the Hexagenia and belong to the genus Ephemera. They are small, 2 - 3mm, dorsoventrally flattened and tended to be found in association with sandy substrates. Their small size means that the early instars are likely to be underrepresented in the samples. Numerical abundances reach maxima of 400 individuals per meter squared at RMS in June and 450 individuals per meter squared at QUITT in July accounting for 10 and 10% of the sample volumes respectively. Overall volumes are considerably lower, approximately 5 percent of the sample volume, but indicate that the nymphs are present throughout the summer and are probably a di- or tri- voltine population of overwintering generations. (Edmunds et al 1971). The Ephemera, too, are nonpredaceous collector-gatherers (Merritt and Cummins 1975).

Order Trichoptera:




The Trichoptera, or caddisflies, are

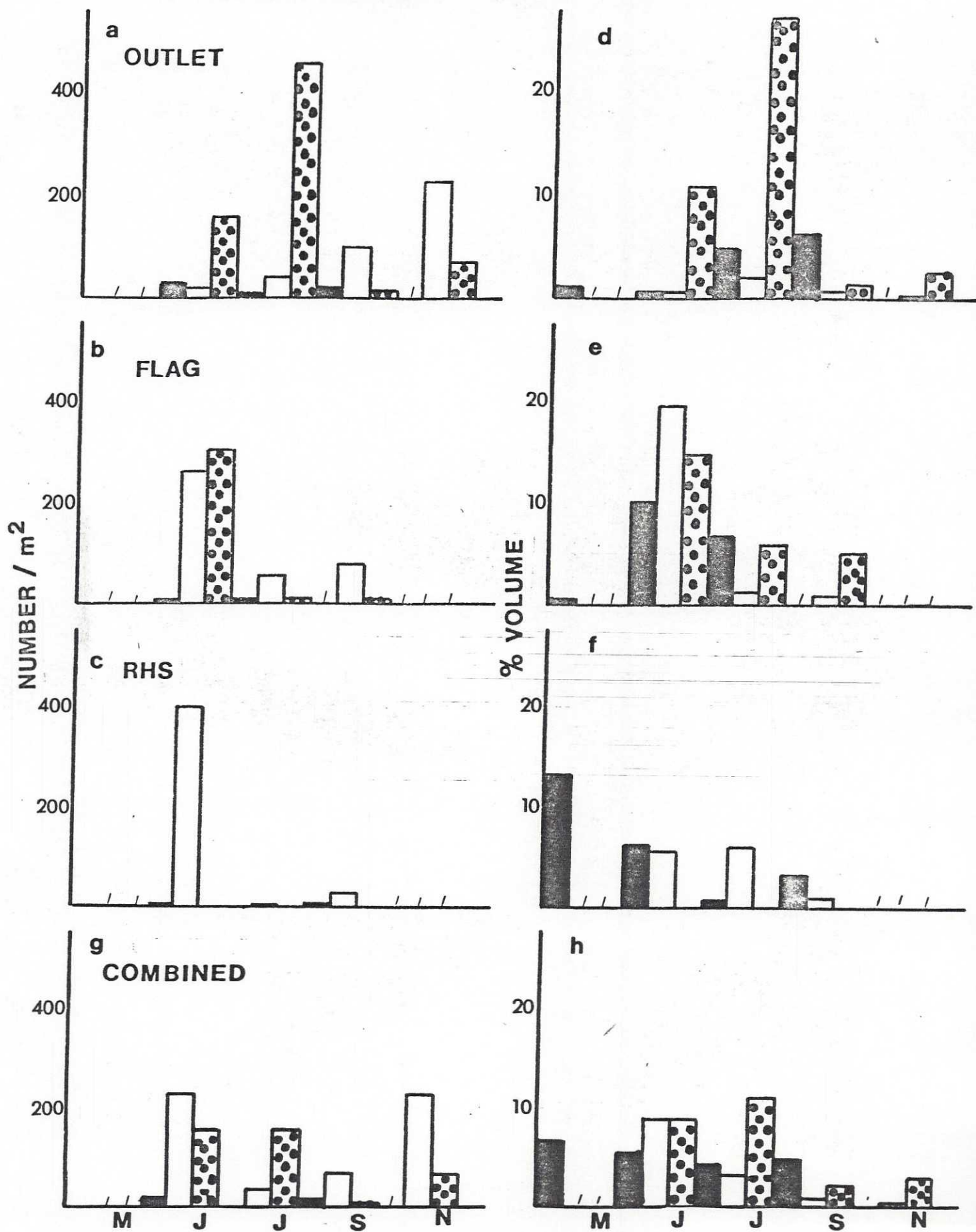
Figure 11:
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f)
contribution of *Sphaerotilus* (Sphaerotopsis) groups to benthic samples from
each of the sites and for all the sites combined (g,h). The different
sampling methods are distinguished (pushnet, shallow water, deep channel).
No samples or deep channel samples were collected during May. No pushnet
samples were collected from the V14C site during September. Only the OUTLET
site was sampled during November.

Figure 11:

Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of Ephemerella (Ephemeroptera) nymphs to benthic samples from each of the sites and for all the sites combined (g,h). The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman). No shallow or deep ekman samples were collected during May. No pushnet samples were collected from the FLAG site during September. Only the OUTLET site was sampled during November.

EPHEMERELLA

 pushnet
 shallow ekman
 deep ekman



holometabolous order of insects with obligate aquatic larvae. The larvae are found in a wide variety of lentic and lotic habitats and are often recognized by the unique tubes or cases they build and often carry with them (Wiggins, 1977). Sunfish Lake trichopteran larvae belong almost exclusively to the family Limniphilidae. The family are non-predatorial collector-gatherers and scrapers (Wiggins, 1977).

A summary of trichopteran larvae in the sample catches is shown in Figure 12. Larvae were invariably found with their cases bound to fronds of Chara. No pupae were caught during sampling. For convenience their size distribution is given in terms of individuals greater or less than 9mm in total length exclusive of cases.

Low population densities persisted at one or more of the sample sites throughout the summer. In the late summer and early fall the numbers of individuals increased to over 370 individuals per meter squared with a consequent 4-fold increase in the percent volume contribution made by the group. The numerical distribution of larvae is clearly associated with sampling habitat indicating a localization to the dense Chara at depths characteristic of shallow ekman samples. The high November population at large size suggests that the

polymorphic order of insects with different aquatic larvae. The larvae are found in a wide variety of aquatic and lotic habitats and are often recognized by the unique shape of cases they build and often carry with them (Wiggins, 1977). Southern Lake Eutrophication larvae belong almost exclusively to the family Limnephilidae. The family are non-predatorial collector-gatherers and scrapers (Wiggins, 1977).

A summary of Eutrophication larvae in the sample catches is shown in Figure 12. Larvae and their cases were found only found with their cases bound to heads of Chironomus. The pupae were caught during sampling. For convenience their size distribution is given in terms of individuals greater or less than 0.5 mm in total length exclusive of cases.

Low population densities persisted at one or more of the sample sites throughout the summer. In the late summer and early fall the numbers of individuals increased to over 100 individuals per meter squared with a consequent 4-fold increase in the percent volume contribution made by the group. The numerical distribution of larvae is clearly associated with sampling method indicating a localization to the dense Chironomus at depths characteristic of shallow stream habitats. The high November population of larvae also suggests that the

Figure 13:

Seasonal changes in the numbers (a,b,c) and the percent values (d,e,f) of *Y. rathbunae* larvae in Pacific samples from each of the sites and for all the sites combined (g,h). Seasonal changes in the numbers of individuals (i,j,k) are shown. The different sampling methods are distinguished.




No shallow or deep shrimp samples were collected during May. No seasonal changes were collected at the TAO site during September. Only the CRITERIUM site was sampled during November.

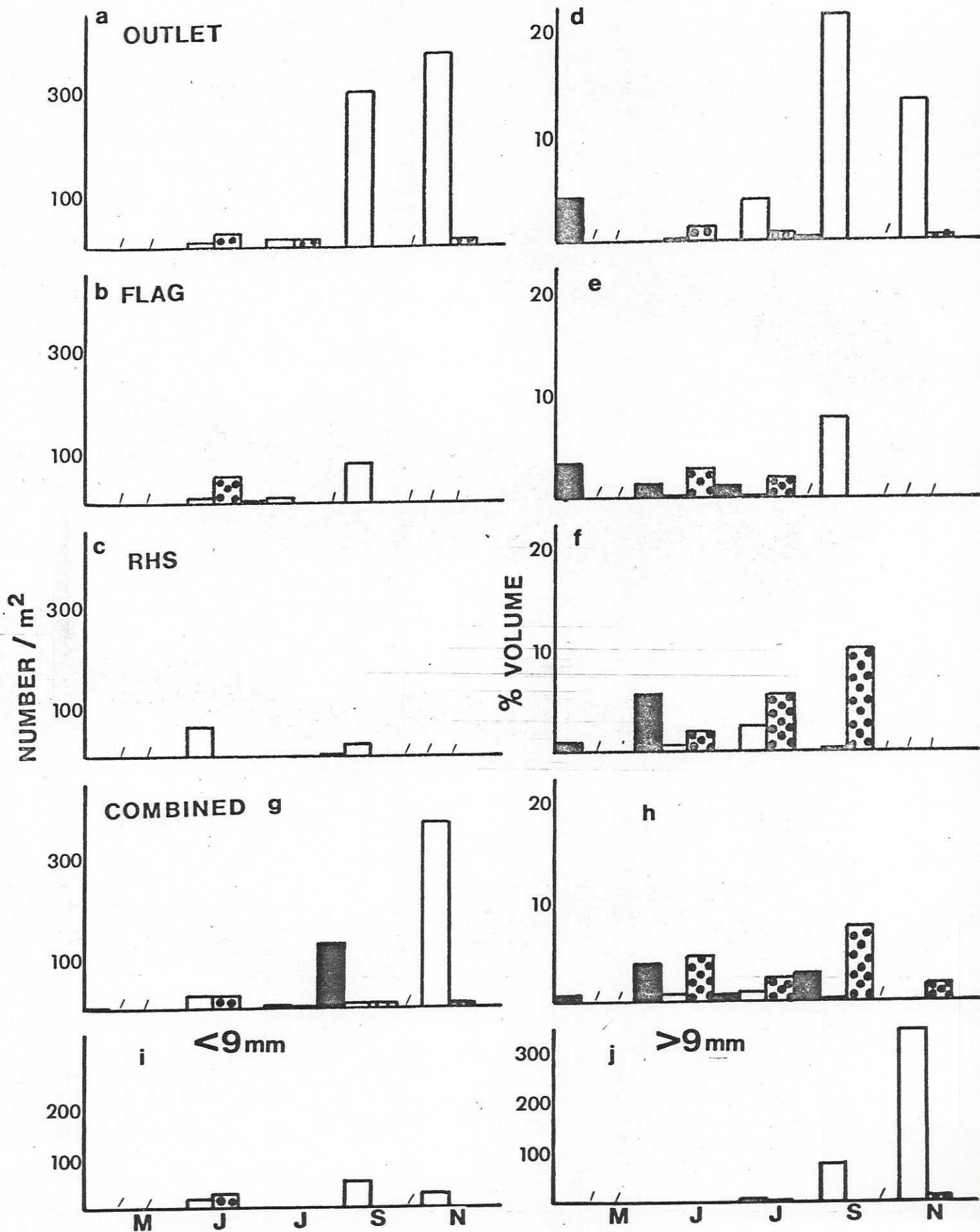
Figure 12:

Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of Trichopteran larvae to benthic samples from each of the sites and for all the sites combined (g,h). Seasonal changes in the numbers of individuals < 9 mm and > 9 mm (i,j) are shown. The different sampling methods are distinguished.

No shallow or deep ekman samples were collected during May. No pushnet samples were collected at the FLAG site during September. Only the OUTLET site was sampled during November.

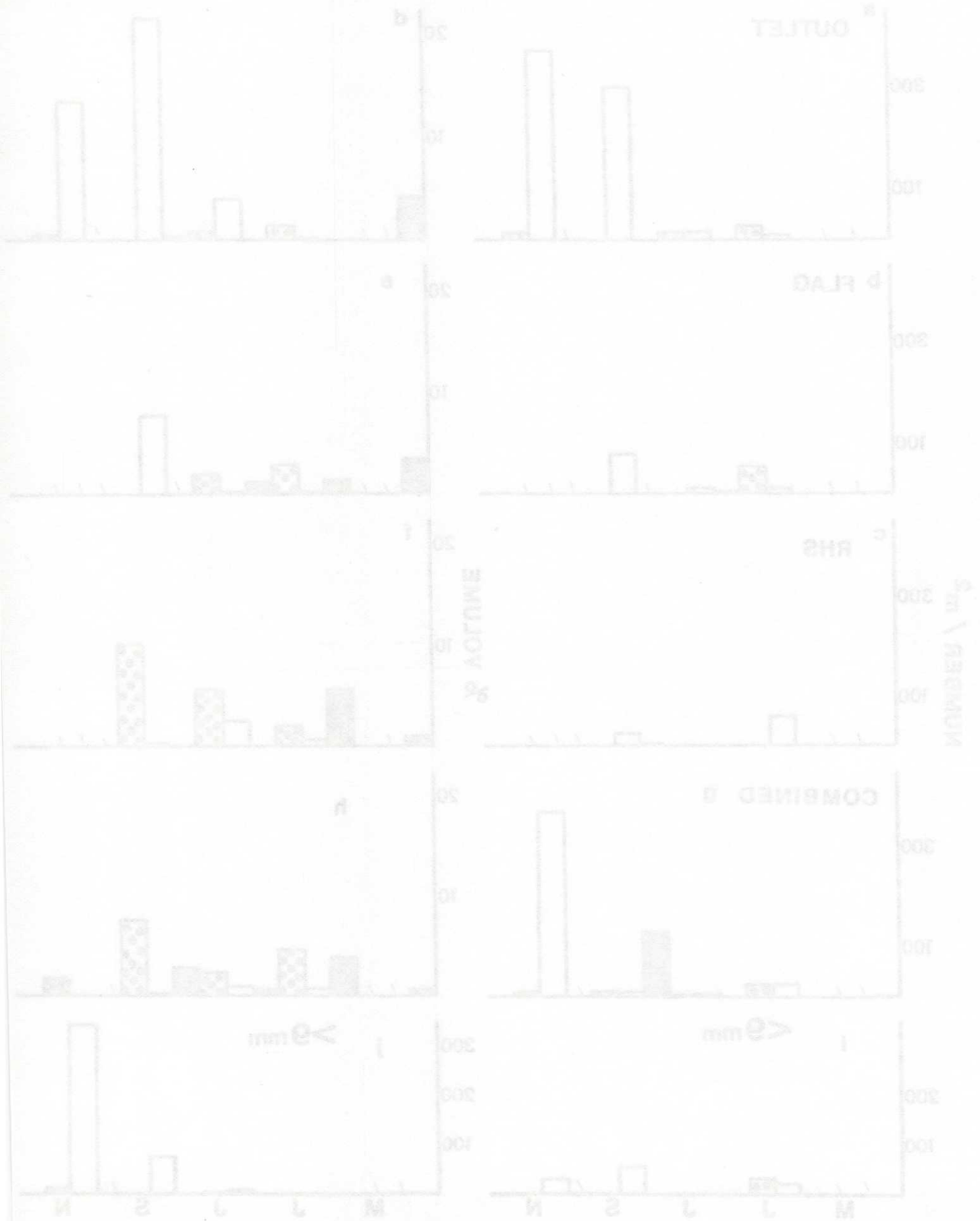
TRICHOPTERA

 pushnet
 shallow ekman
 deep ekman



TRICHOPTERA

Shallow stream
Deep stream



larvae overwinter as sub-adults. This is in accord with other data on the family (Wiggins 1977).

1919-1920. This is in accord with

other data on the family (Wiggins 1977).

Mollusca

The molluscs of Sunfish Lake represent 3 species of gastropods and one species of Lamellibranch which is a rare occurrence. One of the gastropods, Viviparus georgicanicus was present in great abundance and is treated as a single unit, (see Figure 13). The remainder of the molluscs collected are considered together (see Figure 14). The gastropods are grazers of microalgae and microfauna, particularly where epiphytic on the Chara, and also consume benthic detritus.

Gastropoda:

Viviparus georgicanus: This species of gastropod was numerically important at all sites in all months except May. Very low numbers tended to be associated with pushnet samples and the highest levels with shallow ekman samples at the OUTLET site. There is a tendency to move to depth in November presumably to overwinter. This overwintering at depth is probably the cause of the small numbers collected in May. Also shown in Figure 13 are the size distributions of the Viviparus based on the range of 1 to 6mm in maximum length. The June distribution is broad relative to the distribution in other months, particularly September, when the OUTLET site shows a considerable recruitment of smaller individuals.

The following 7 species represent a species of gastropods and one species of bivalves which is a rare occurrence. One of the gastropods, *Viviparus* *neogibbatus* was present in great abundance and is treated as a single unit. (see Figure 11). The remainder of the molluscs collected are considered together (see Figure 11). The gastropods are present in micaceous and micaceous, particularly when ephaptic on the *Chama*, and also common bivalve dentures.

Gastropods:

Viviparus neogibbatus. This species of gastropod was numerically important at all sites in all months except May. Very few numbers tended to be associated with banded samples and the highest levels with shallow ocean samples at the OUTLET site. There is a tendency to move to depth in November presumably to overwinter. This overwintering at depth is probably the cause of the small numbers collected in May. Also shown in Figure 11 are the size distributions of the *Viviparus* based on the range of 1 to 6mm in maximum length. The June distribution is most relative to the distribution in other months, particularly September, when the OUTLET site shows a considerable recruitment of smaller individuals.

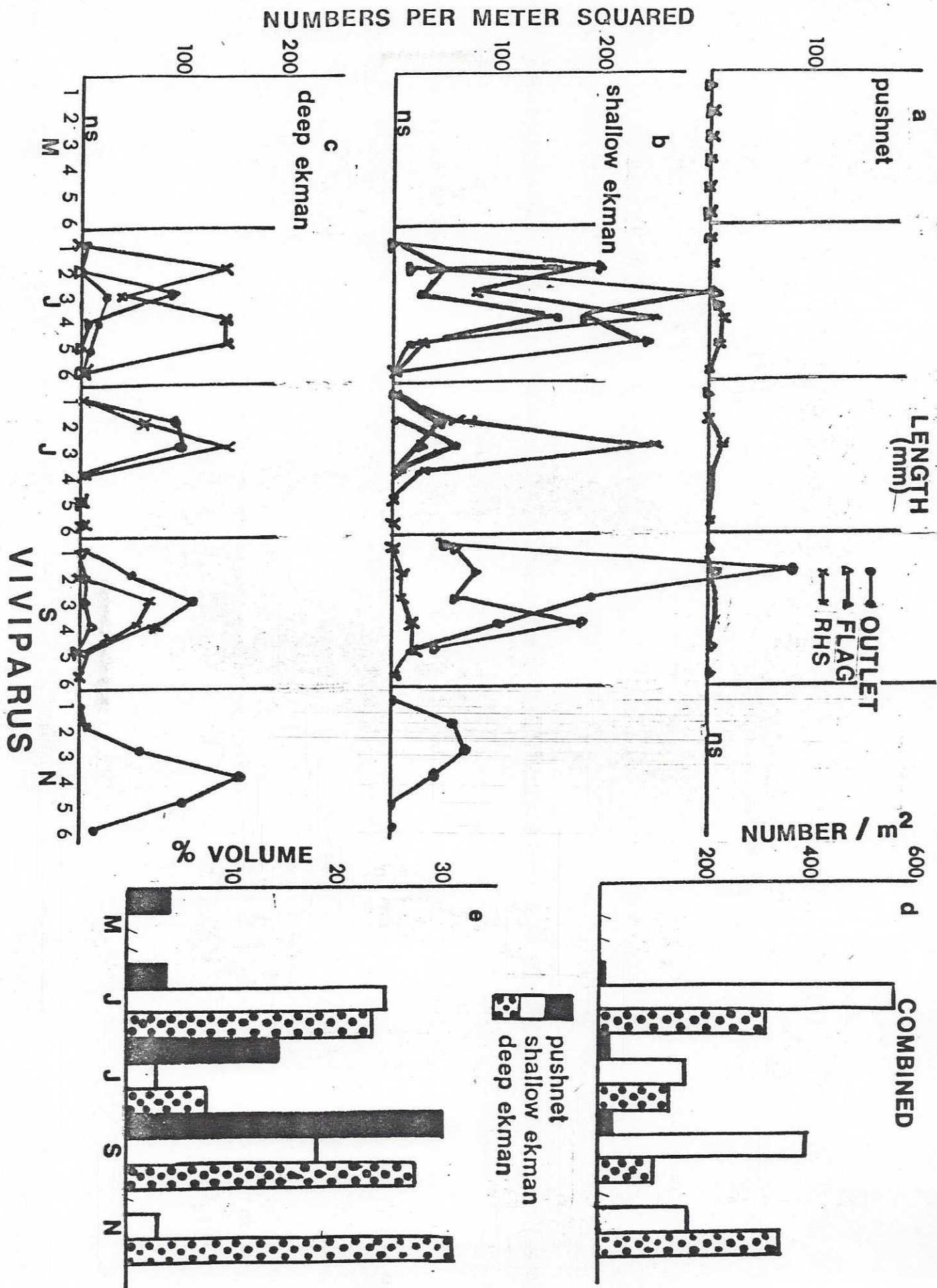
Figure 11:
Seasonal changes in the size distribution of *Viviparus georgianus*
(Gastropoda) are shown by the sampling method used (a,b,c). The different
sites are distinguished (OUTLET, PLAC, MAY).
Seasonal changes in the numbers (d) and the percent volume (e) composition
of *Viviparus* to the beachic species are shown for all the sites combined.
The different sampling methods are distinguished (pushnet, shallow sump,
deep sump).
No shallow or deep sump samples were collected during May. No pushnet
samples were collected from the PLAC site during September. Only the OUTLET
site was sampled during November.

Figure 13:

Seasonal changes in the size distribution of Viviparus georgianicus (Gastropoda) are shown by the sampling method used (a,b,c). The different sites are distinguished (OUTLET, FLAG, RHS).

Seasonal changes in the numbers (d) and the percent volume (e) contribution of Viviparus to the benthic samples are shown for all the sites combined. The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman).

No shallow or deep ekman samples were collected during May. No pushnet samples were collected from the FLAG site during September. Only the OUTLET site was sampled during November.

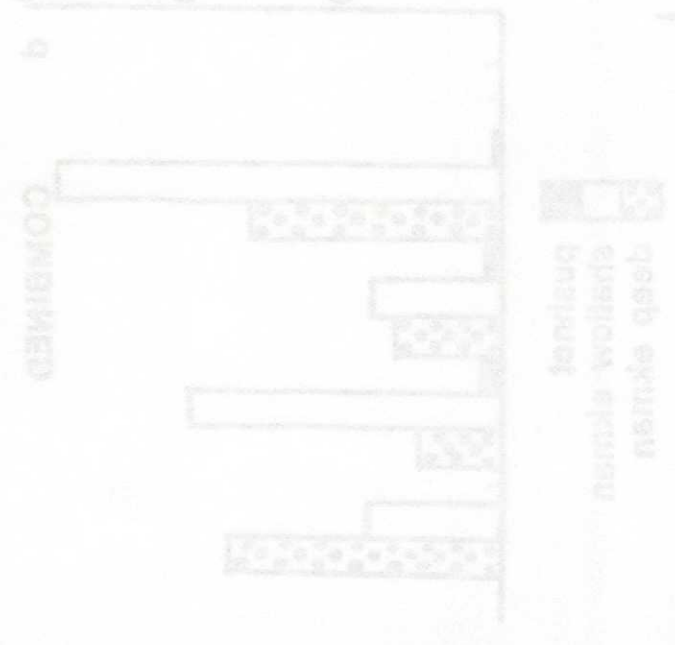
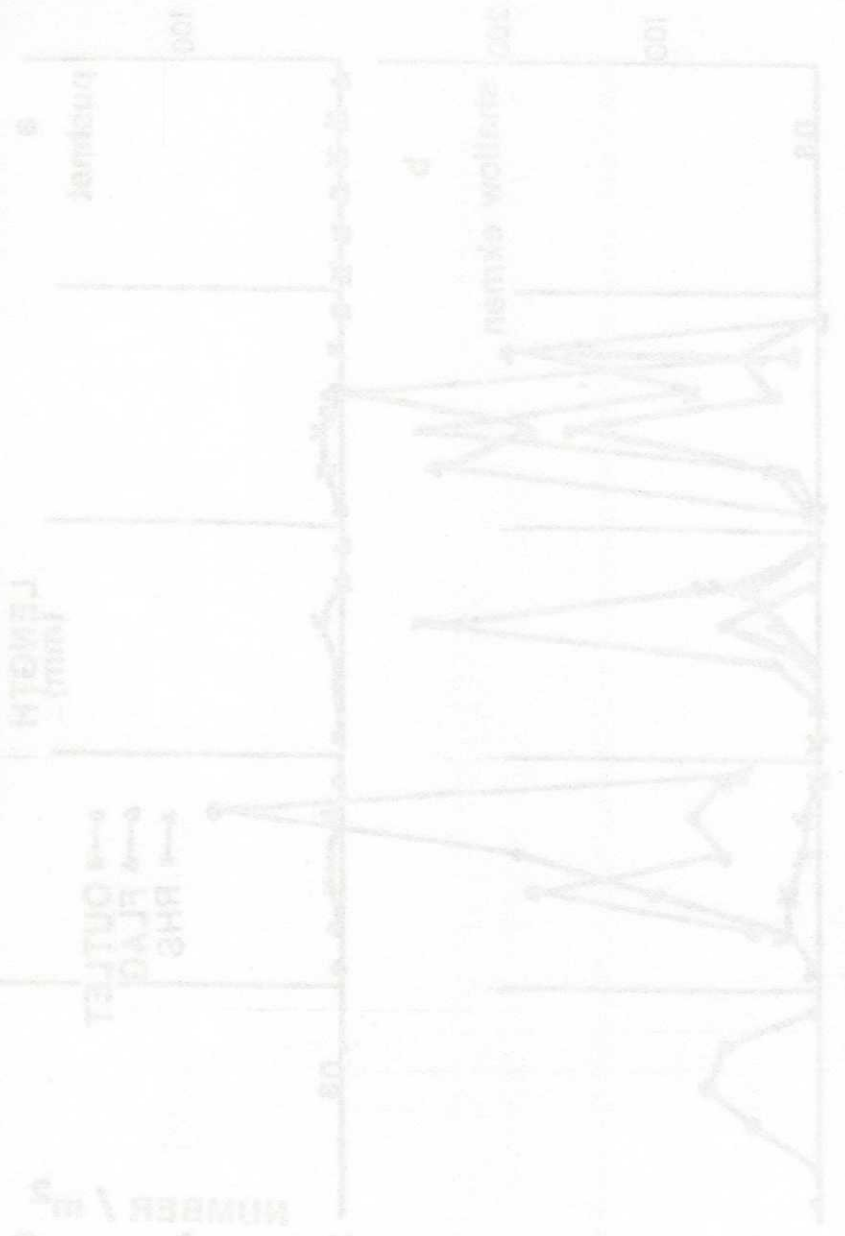
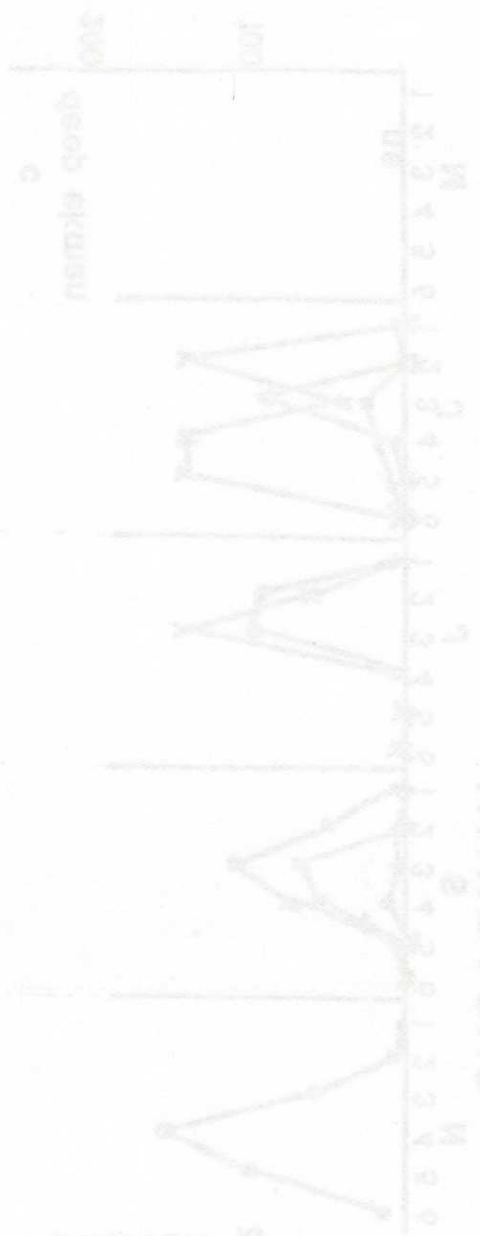


2

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2

VIVIPARUS



The absence of larger individuals late in the summer indicates that the species may be annual (see also Turnbull, 1975) overwintering at a mean size of 4mm. Particularly at depth the Viviparus account for a relatively large proportion of the percent volume in samples. Several of the 3.5 meter samples without Chara contained only Viviparus and Chironomids.

Other molluscs:

The other molluscs are dominated by Physa, a large bodied gastropod found exclusively on Chara. A slight contribution to the numbers and percent volume is made by Gyraulus, a dorsoventrally flattened gastropod whose shells are common in the sediments but of which less than 10 live individuals were collected in the course of sampling. The final contribution to the molluscan fauna is Pisidium, a small, (6-8mm) white bivalve collected alive only 3 times. The Physa are numerically important at the FLAG sample site in July although they compose only 6 percent of the volume at that time. The largest volume contributions by the Physa are made in low diversity samples where their individual bulk becomes important relative to that of small bodied Chironomids (pers. obs).

Crustacea:

Order Amphipoda:

The abundance of larger individuals late in the summer indicates that the species may be annual (see also Forman, 1972) overwintering at a mean size of 1mm. Particularly at depth the Viviparus account for a relatively large proportion of the percent volume in samples. Several of the 2.5 meter samples without Chama contained only Viviparus and Chironomids.

Other molluscs:

The other molluscs are dominated by Physa, a large bodied gastropod found exclusively on Chama. A slight contribution to the numbers and percent volume is made by Gyrinus, a dorsoventrally flattened gastropod whose shells are common in the sediments but of which less than 1% live individuals were collected in the course of sampling. The total contribution to the molluscan fauna is Physa, a small (15-25mm) white bivalve collected alive only 3 times. The Physa are numerically important at the ELAC sample site in July although they compose only 1.6 percent of the volume at that time. The largest volume contributions by the Physa are made in low diversity samples where their individual weight becomes important relative to that of small bodied Chironomids (pers. obs.).

Crustaceans:

Order Amphipoda:

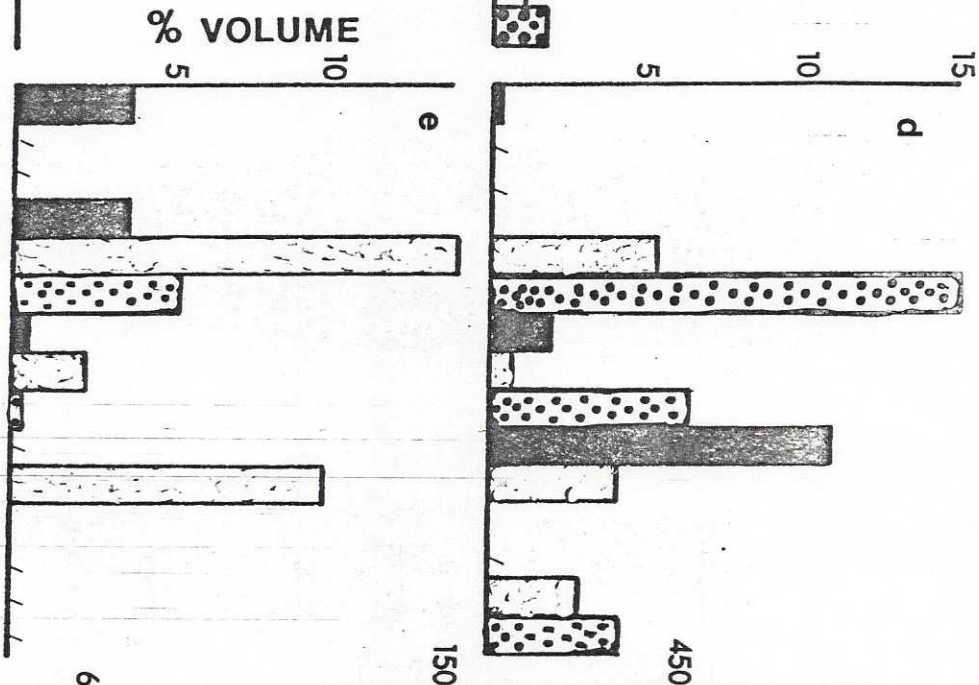
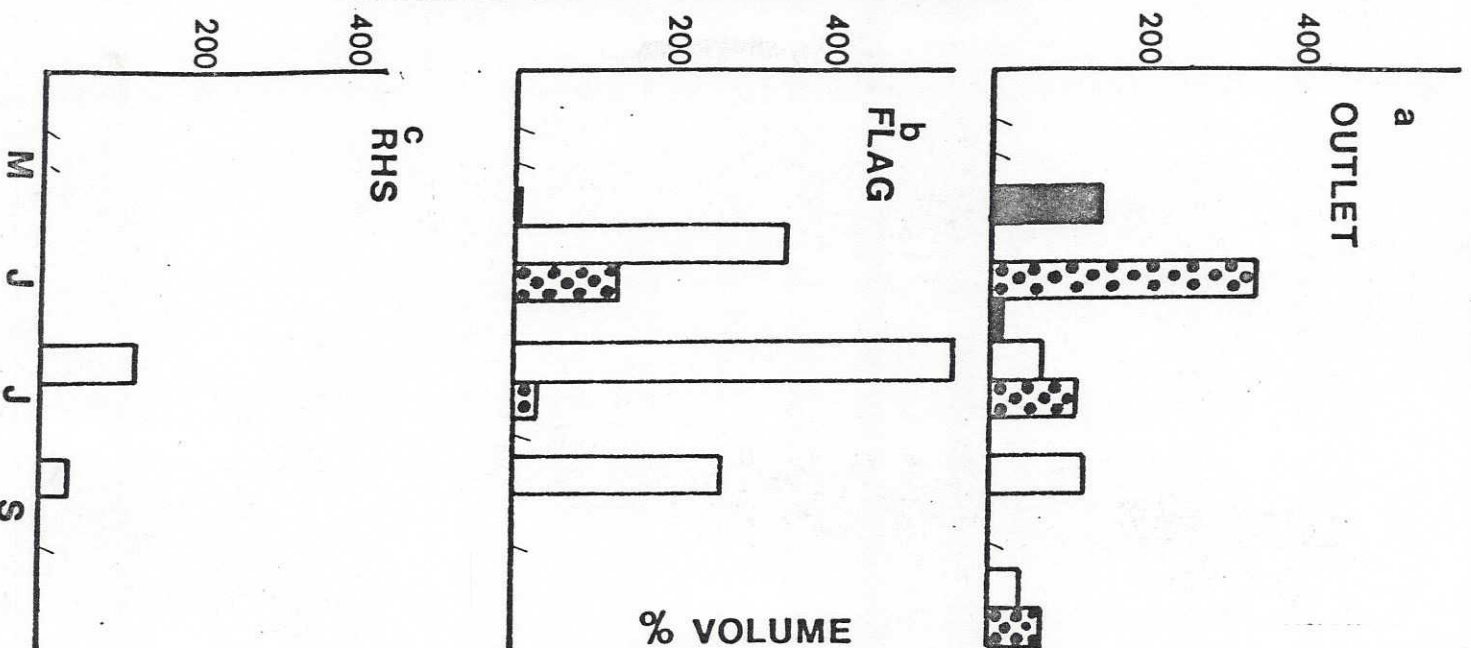
Figure 14:
Seasonal changes in the numbers (a, b, c) and the percent volume (d, e, f)
composition of other bacteria (predominantly *Escherichia coli*) in benthic samples
from each of the sampling sites and for all the sites combined (g, h).
The different sampling methods are distinguished (quadrat, shallow drag,
deep drag).
No station or deep drag samples were collected during May. No percent
samples were collected from the T10 site during September. Only the *Escherichia coli*
size was sampled during November.

Figure 14:

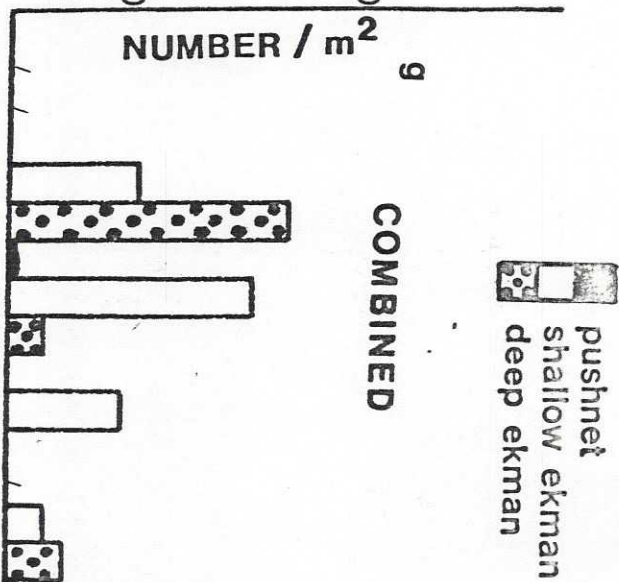
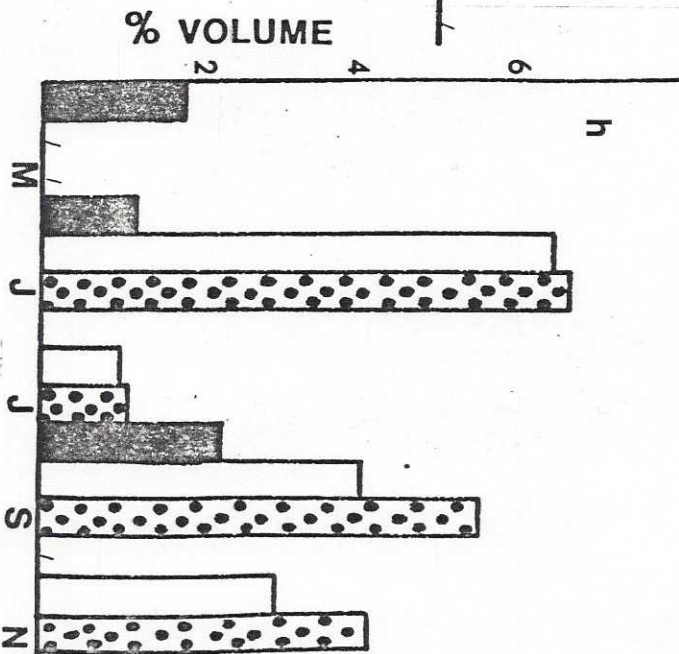
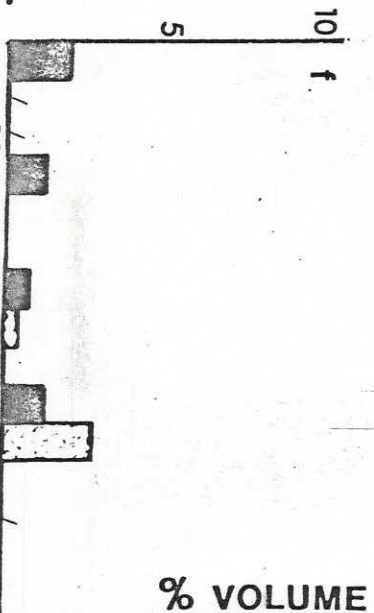
Seasonal changes in the numbers (a,b,c) and the percent volume (d,e,f) contribution of other Gastropods (predominantly Physa)to benthic samples from each of the sampling sites and for all the sites combined (g,h). The different sampling methods are distinguished (pushnet, shallow ekman, deep ekman).

No shallow or deep ekman samples were collected during May. No pushnet samples were collected from the FLAG site during September. Only the OUTLET site was sampled during November.

NUMBERS PER METER SQUARED

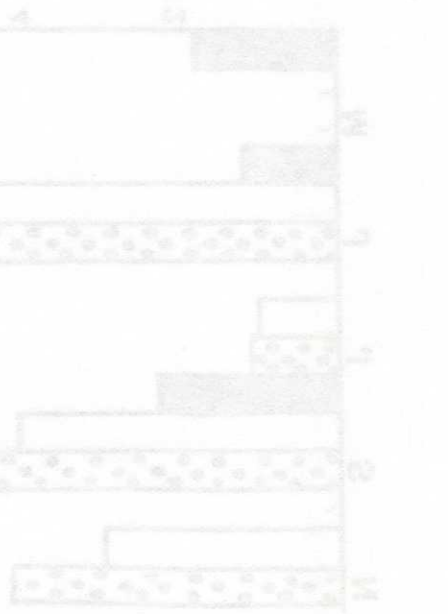


OTHER GASTROPODS



pushnet
shallow ekman
deep ekman

OTHER CASTROPODS



NUMBERS PER METER SQUARED



FLAG

% VOLUME

% VOLUME

OUTLET

COMBINED

deep exman
shallow exman
combined

The amphipod population of Sunfish Lake is monospecific, composed of Hyalella azteca individuals. Figure 15 shows the size distribution of Amphipods in the samples. The intermediate (2 - 3mm) sized individuals predominate throughout the season. In June there are appreciable numbers of the largest individuals (>3mm) which presumably constitute the breeding population (Cooper, 1965). Throughout the remainder of the season, the intermediate (2 - 3mm) individuals predominate. A substantial recruitment of small individuals is seen in July and again in September. By November, reproduction has ceased and the distribution becomes far more compact, consisting of the overwintering population of adults and subadults (Cooper, 1965). High numerical abundances and percent volumes are clearly associated with shallow samples and the protection afforded by the Chara. Amphipods are a reliable indicator of the presence of Chara in a sample (see Table H). The overwintering population constitutes a 3 fold increase over the observed September values indicating growth of a September recruitment of individuals to a size detectable by the sampling methods used. Seasonal variation in the abundance of amphipods is very large, with a November surface ekman mean of 4250 individuals per meter squared and a June surface ekman mean of less than 100 individuals per meter squared.

The sampled population of *Amphipoda* is monospecific, composed of *Hyalella* species individuals. Figure 1 shows the size distribution of *Amphipoda* in the samples. The intermediate (2 - 3mm) sized individuals predominate throughout the season. In June there are appreciable numbers of the largest individuals (4mm) which presumably constitute the breeding population. Throughout the remainder of the season, the intermediate (2 - 3mm) individuals predominate. A substantial recruitment of small individuals is seen in July and again in September. By November, reproduction has ceased and the distribution becomes far more compact, consisting of the overwintering population of adults and subadults (Cooper, 1965). High numerical abundances and percent volumes are directly associated with shallow samples and the production allowed by the Chert. *Amphipoda* are a reliable indicator of the presence of Chert in a sample (see Table 2). The overwintering population constitutes a fold increase over the observed September values indicating growth of a September recruitment of individuals to a size detectable by the sampling methods used. Seasonal variation in the abundance of *Amphipoda* is very large, with a November surface count mean of 1250 individuals per meter squared and a June surface count mean of less than 100 individuals per meter squared.

Figure 13:

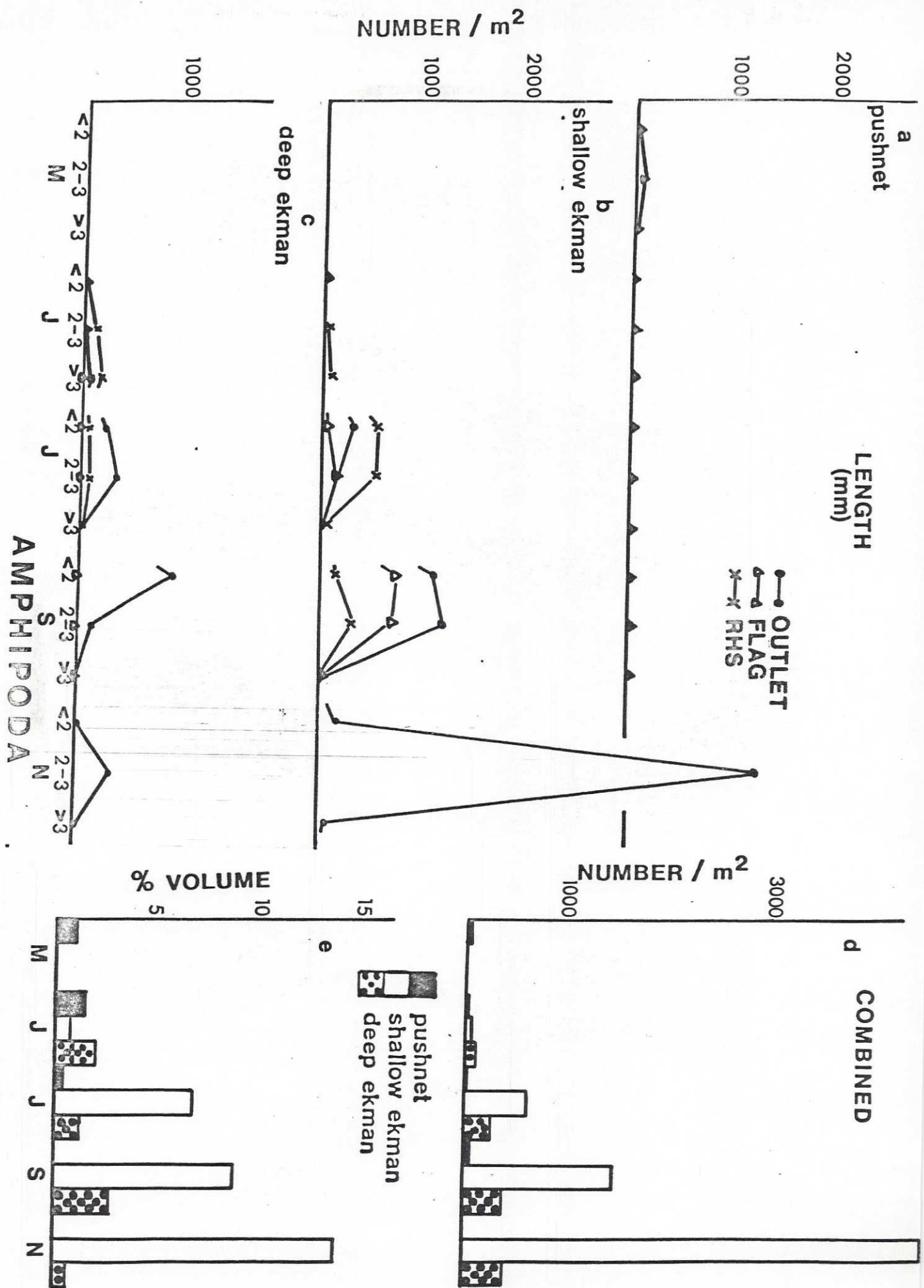
Seasonal changes in the size distribution of *Hyalella* species (Amphipoda) are presented by the sampling method used (a, b, c). The different sampling sites are distinguished (GOLLET, PAC, NAB). Seasonal changes in the numbers (d) and the percent volume (e) composition are presented for all the sites combined.

No shallow or deep clean samples were collected during May. No further samples were collected from the PAC site during September. Only the GOLLET site was sampled during November.

Figure 15:

Seasonal changes in the size distribution of Hyaella azteca (Amphipoda) are presented by the sampling method used (a,b,c). The different sampling sites are distinguished (OUTLET, FLAG, RHS). Seasonal changes in the numbers (d) and the percent volume (e) contribution are presented for all the sites combined.

No shallow or deep ekman samples were collected during May. No pushnet samples were collected from the FLAG site during September. Only the OUTLET site was sampled during November.



2

3

4

AMPHIBODA



Hydracarina:

The mite population is dominated by a soft-bodied, red mite that lives exclusively in association with Chara. During their peak abundances in July and September the mites were visible from the water surface as bright red mottlings of the Chara fronds. Minimal numbers (a maximum of 40 individuals per meter squared) were found of a group of hard bodied mites. These were distinguished in the collections but all the mites are presented collectively in Figure 16. The mites are small relative to other benthic macroinvertebrates studied (1 - 2.5mm) so they never contribute more than 4% of the sample volume although the numerical abundance reaches 210 individuals per meter squared in surface ekman samples at the FLAG site during September. The mites are virtually absent from May and June samples, thereafter they increase rapidly in abundance. It is probable that the group is univoltine and that smaller individuals or larvae were not retained in June. Little is known of the natural history of the Hydracarina (Parker, 1975).

Phylum Annelida:

Oligochaeta

The mite population is dominated by a soft-bodied
red mite that lives exclusively in association with
Chiron. During their peak abundance in July and
September the mites were visible from the water surface
as bright red mottlings of the Chiron. Mites
numbers a maximum of 40 individuals per water spider
were found of a group of hard-bodied mites. These were
distinguished in the collections but all the mites are
presented collectively in Figure 16. The mites are
small relative to other aquatic macroinvertebrates
studied (1 - 1.5mm) so they never contributed more than
4% of the sample volume although the numerical abundance
reaches 118 individuals per water spider in some
stagnant samples at the F&B site during September. The
mites were virtually absent from May and June samples,
characterized by increase rapidly in abundance. It is
probable that the group is univoltine and that smaller
individuals or larvae were not retained in June. Little
is known of the natural history of the Hydracarina
(Parker, 1973).